



Pathway A: traditional nucleation Pathway B: via amorphous precursor (PhosphoProtein C-terminal) Königsberer E. et al. 2006

Non-hierarchical: Direct nucleation Pathway C: additives stabilises ACP Hierarchical: single crystals without planes . Liu Y. et al. 2011.



Experimental Design



The ability of a nanobioglass-doped self-etching adhesive to re-mineralize and bond to artificially demineralized dentin

Abuna, Gabriel¹⁻³. Campos, Ferreira Paulo¹. Hirashi, Noriko². Giannini, Marcelo¹. Nikaido, Toru². Tagami, Junji². Geraldeli, Saulo ³. Sinhoreti, Coelho Mario Alexandre¹ 1. Restorative Dentistry Department, Dental Materials Division, Piracicaba Dental School, State University of Campinas, Piracicaba, Sao Paulo, Brazil. 2. Cariology and Operative Dentistry Department, Tokyo Medical and Dental University, Tokyo, Japan. 3. General Dentistry Department, Division of Biomedical Materials, East Carolina University School of Dental Medicine, Greenville, USA.

Research Question: Can we create a nanoparticle able to remineralize Caries affected Dentin, and improve the longevity of the dental restorations through time?



TEM Mode: Imaging

50 nm HV=75.0kV Direct Mag: 200000x AMT Camera System

Print Mag: 181000x @ 7.0 in 12:33:33 11/28/2017 TEM Mode: Imaging

AMT Camera System

100 nm HV=100.0kV Direct Mag: 100000x AMT Camera System

TEM Mode: Imaging

24 hr	1month													
			1.2 -											
	1 -	1												
			0.8		``	``								
			0.6											
			0.4 -	0.4										
			0.2											
			0 -						` ~ .					
20 30 40 50 60 70 80 90 100			C) 10	20	30	40	50	60	70	80	90	100	
– – experimenta														
Clearfill			Experimental				Nanoparticle							
24hr	1month	24hr		1month		24hr		1	1month					
74.0	78.8	81.3		73.8		57.0				78.1				
50.4	53.7	37.4			38.3			42.4			58.9			
34.5	36.6	17.3		19.9			31.6				44.5			
01 /	00.0	0	0		0.0		01.0				01.0			



We assess the recover of mechanical property of the tissue affected, it surely recover the hardness, improving also the Elastic modulus and the stiffness. Properties directly related with the longevity of the restoration and the health of the tissue.

The Weibull m, provide a perception of how the material will behave in hypothetical time through mathematical calculus. It was calculated from the µTBS that measure the bonding strength of the material bonded to the tissue.





experimental24hr nanopt24hr



Fisher LSD, p<0.05







- \mathcal{R}
- http://dx.doi.org/10.1038/nmat2875

- Θ

References

1. Boskey AL, Roy R. Cell culture systems for studies of bone and tooth mineralization. Chemical Reviews. 2008;108(11):4716-

2. Cölfen H. Biomineralization: A crystal-clear view. Nature materials [Internet]. Nature Publishing Group; 2010 Dec [cited 2013] Dec 16];9(12):960–1. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21102512

3. Nudelman F, Pieterse K, George A, Bomans PHH, Friedrich H, Brylka LJ, et al. The role of collagen in bone apatite formation in the presence of hydroxyapatite nucleation inhibitors. Nature Publishing Group; 2010;9(October):9–14. Available from:

4. Abuna G, Feitosa VP, Correr AB, Cama G, Giannini M, Sinhoreti MA, et al. Bonding performance of experimental bioactive/biomimetic self-etch adhesives doped with calcium-phosphate fillers and biomimetic analogs of phosphoproteins. Journal of Dentistry [Internet]. Elsevier Ltd; 2016;52:79–86. Available from: http://linkinghub.elsevier.com/retrieve/pii/S0300571216301439

5. Amos FF, Olszta MJ, Khan SR, Gower LB. Relevance of a Polymer-Induced Liquid-Precursor (PILP) Mineralization Process to Normal and Pathological Biomineralization. Biomineralization - Medical Aspects of Solubility. 2007. 125-217 p. 6. Märten A, Fratzl P, Paris O, Zaslansky P. On the mineral in collagen of human crown dentine. Biomaterials. 2010;31(20):5479-

7. Kokubo T, Takadama H. How useful is SBF in predicting in vivo bone bioactivity? Biomaterials. 2006;27(15):2907–15. 8. Tsuda H. Raman spectra of human dentin mineral. European Journal of Oral Sciences. 1996;73(2):1703–131.